Enzymatic Resolution of Aziridine-Carboxylates.

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Abstract: N-acyl-aziridine-2-carboxylates and N-acyl-aziridine-2,3-dicarboxylates have been resolved with good to excellent stereochemical purity by enzymatic hydrolysis catalyzed by lipase from *Candida cylindracea.*

In recent years, aziridine-2-carboxylates have played an interesting role as chiral synthons in the synthesis of α - and β -aminoacids¹ and β -lactams;² the development of new and easy ways of synthesizing optically active aziridine-carboxylates has therefore become a matter of great importance. There are relatively **few** generallyapplicable methods available in the literature, 3 and it was in an attempt to find an alternative to chemical methods that we recently discovered⁴ that hydrolases can be efficiently used to resolve three-membered ring heterocycles containing ester groups on the ring carbon atom, such as oxaziridines and N-chloroaziridines. Previous results showed that several hydrolases are quite active in aziridine enantiomer differentiation;⁴ among these, a-chymotrypsin and lipases from *Rhizopus delemar* **and** *Candida cylindrucea* display higher enantioselectivity towards N-H and N-substituted aziridine -carboxylates.⁵

In this paper we report the resolution of racemic N-acyl-2-methoxycarbonylaziridines **(1)** and N-acyl-2,3 bismethoxycarbonylaziridines (2) by enantioselective enzymatic hydrolysis catalyzed by lipase from *Candida cyfindfacea* (CCL).

Racemic aziridines **1** and 2 were prepared by acylation of the corresponding N-H aziridines with acetyl or butyryl chloride, as described in the literature.⁶ CCL was purchased from Sigma and used without purification.

Hydrolyses were performed in phosphate buffer $(pH 7.5; 0.1 M)$, standard conditions which we have already used advantageously for the selective resolution of other substrates,⁴ and transformations were usually stopped at 60% of conversion. Under these conditions, CCL catalyzed the hydrolysis of the aziridines **1 and 2**

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with **high enantioselectivity and chemoselectivity; surprisingly, however, it showed a very high esterolytic activity towards substrate 2, but primarily catalyzed the hydrolysis of the amide bond in compounds 1. Enzymatic hydrolysis of racemic derivatives** 1, **up to 50% conversion, thus afforded the unhydrolyzed esters** 1 and the corresponding aziridine 3 as enzymatic hydrolysis product, both with good to excellent stereochemical **purity (Table 1, Scheme 1).**

 $a)$ $R = CH_3$; b) $R = n \cdot C_3H_7$

The optically-active unreacted esters **1** and the aziridine 3 were isolated from the aqueous phase by simple extraction with CH_2Cl_2 . Owing to its high volatility, aziridine 3 was recovered by conversion into the corresponding N-chloro derivative whose absolute configuration and optical purity are well established.7 After extraction, the combined organic layers were therefore treated with t-butylhypochlorite⁴ at -10 °C, then concentrated and the residue was chromatographed on silica gel (CH2C12/ethyl ether as eluant). The recovered unchanged aziridines **la and lb** showed 63% and 90% enantiomeric excess (ee.), respectively. It is worth noting that, while higher conversion did not improve the optical purity of **la,** stopping the reaction at 20% conversion afforded the aziridine 3 in 80% ee.

With respect to compounds 2, enzyme-catalyzed hydrolysis provided the optically-pure unreacted esters 2 and the corresponding N-acyl-2,3-dicarboxylic acid monomethylesters 4 (Scheme 1). The unhydrolyzed esters 2 **were isolated from the reaction mixture by extraction with CH2C12 followed by column chromatography on**

silica gel $\left(\text{CH}_2\text{Cl}_2/\text{ethyl} \right)$ ether as eluant). The enzymatic-hydrolysis products 4 were recovered from the aqueous phase by acidification with HCI 10% and extraction with ethyl ether. The optical purities of monoesters 4 were estimated through their conversion into the corresponding diestcrs 2 by esterification with diazomethane. The absolute configuration of aziridines 2 was determined by its conversion into the free NHdiacid by hydrolysis with lithium hydroxide.⁸

Substrate	Reaction condition			unchanged aziridine ^a			
	t/h	E/S ^b	conversion% ^c	vield%	$[\alpha]_D$ ^d	$ee%$ ^e	conf.
1a	4	1/4	55	20	-46.1	63	2S ^f
1b	1	1/20	60	35	-74.3	90	2S ₈
2a	24	1/1	60	30	-56.4	295	2R,3R ^h
2b	3	1/1	70	25	-34.8	295	2R,3R ^h

Table 1. Enzymatic hydrolysis of aziridines 1 (a,b) and 2 (a,b) catalyzed by lipase from *Candida cylindracea*.

a) Nmr properties and mass spectra are consistent with the structures considered. b) All hydrolyses were performed in phosphate buffer (PH 7.5: 0.1 M) at room temperature. Reaction mixture defined from the ratio (w/w) enzyme/aziridine (E/S). c) Conversion calculated from the relationship 11 c = ee_s/ee_s+ee_n where ee_s (enantiomeric excess of substrate) refers to the recovered unchanged aziridines and $ee₀$ (enantiomeric excess of product) refers to the hydrolyzed product. The hydrolyzed product 3 showed 40% and 60% ee, from **la** and **lb** respectively, evaluated as N-chloro derivatives. The monoesters 4 showed 60% and 30% ee, from **2a** and 2b, respectively. d) Data from chloroform solution. e) $\pm 2\%$. Optical yield (ee%) determined from ¹H nmr spectra recorded in CDC13 solution and in presence of Eu(hfc) $_3$. f) from ref. 6. g) Assigned by acylation of the hydrolyzed product (2R)-(+)-3.7 h) assigned by correlation with diethyl aziridine-2,3-carboxylate.⁸

The results reported in Table 1 show that the CCL-catalyzed hydrolysis of aziridines 1 and 2 affords good yield with excellent enantioselectivity. N-acetyl aziridines **la** and **2a were** hydrolyzed more slowly than the corresponding butyryl **lb** and **2b;** moreover, the change of the N-substituent from acetyl to butyryl in aziridine-2-carboxylates significantly increased enantioselectivity. In our opimon, this is a very easy and efficient way of obtaining both enautiomers of aziridine-carboxylates in good optical and chemical yield. The synthetic potential of these results is strengthened by the nature of the chiral aziridines **1** and 2: both are activated towards nucleophilic ring opening reactions and therefore afford easy access to functionalized aminoacid-derivatives.I,9.lo

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